



Influence of sodium alginate on growth and elements content of *Corchorus olitorius*, *Vicia faba* & *Zea mays* plants as well as soil properties.

Rasha Mohamed Eid Gamil, Samia Ali Haroun, Aamaa Omar Hegazy

Received: 6/2/2020
Accepted: 29/2/2020

Abstract: Sodium alginate (SA) has proved as a plant growth promoting substance for various medicinal and agricultural crops. A pot experiment was carried out to explore the effect of different SA concentrations (25, 50 & 100 ppm) on plant growth and physiological activities in *Corchorus olitorius*, *Vicia faba* and *Zea mays* after 10, 20 and 40 days from sowing. Presoaking in the used SA concentrations significantly improved the performance of the tested plants. Results demonstrated that the used SA concentrations significantly increased growth parameters (root length, shoot length, fresh & dry weight, and water percentage/ plant), soil physical and chemical properties and the determined elements (N, P, K and Na) especially 50 ppm which caused the best influences.

keywords: alginate, growth, elements & soil.

1. Introduction

Nowadays there is an increase in the world population, especially developing countries as Egypt so, there is an urgent need for food especially that of plant origin which should be safe for human consumption or animal that used for human being. There are many substances used as growth stimulators to increase the growth, density and productivity of the plant through the activation of vegetative growth and vital processes, but some of them are harmful and dangerous to human health so, it was necessary to search for the safe ones for humans and animals such as sodium alginate.

Alginates possess very important position among the natural polysaccharides considered as structural part of the brown algae *Sargassum* [5] in very huge amounts. Sodium alginate (SA), a recent multifunctional marine bioactive substance derived from the brown algae, consists of homopolymeric poly- β -(1,4) D-mannuronic acid residue and poly- α -(1,4) L-gluronic acid residues. Sodium alginate in its depolymerized form, known as oligo-alginate, is prepared by hydrolysis of acid or degradation of enzyme of sodium alginate. Oligomers with comparatively low molecular weight. Application of these oligomers on plants results in different vital and physiological activities, including growth promotion of plant generally, germination of seed, elongation of shoot, root development, production of flower,

antimicrobial activity, amelioration of heavy metal stress, phytoalexin induction, etc. [17, 28, 18, 16, 1, 20].

Corchorus olitorius may be a vegetable, commonly known as "Jew's mallow", "tossa jute", "bush okra", "krinkrin", "molokhia", "West African sorrel" and "jute mallow", it contains large amount of vitamins, minerals and other factors used in nutrition [7]. This is often one of the most consumed widely conventional vegetables in Africa [37] and is also consumed in Asia [24]. Obtained from both wild and cultivated sources. The leaves are consumed as a component of vegetable-based dish that go with starchy dishes. It may be arranged alone or in blends with vegetable takes off (East Africa) or other wild vegetables or in blend enhanced with fish sauce. In this way it plays an amazingly critical part in nourishment security and the fight against destitution in Africa [6].

One of versatile crop is *Vicia faba* (faba bean). It belongs to the Fabaceae family and has the ability to develop in different climates [50]. Besides, it can be utilized around the year, as it can be consumed in both processed and raw forms. The seed grain mostly consumed in the human diet, while the pods are used as feed. However, as the pods also provide micro-, macro and non-nutrient phytochemicals, they

have potential to be utilized as a source of functional compounds [34]. The nutritional importance of faba bean is prominent, approximately 250 g protein/kg seed [33] and it gives a valuable amount of energy: 320 kcal/100 g dry weight [40].

Maize (*Zea mays*), called corn in some positions of the world, belongs to graminaceous plants of tropical origin that has become the main grain crop in the world in terms of total production [36]. About 45–50% of oil that is used in cooking, salads and is obtained from wet milling process comes from maize germs [41]. The oil contains 14% saturated fatty acids, 30% monounsaturated fatty acids, and 56% polyunsaturated fatty acids. The refined maize oil contains linoleic acid 54–60%, oleic acid 25–31%, palmitic acid 11–13%, stearic acid 2–3% and linolenic acid 1% [12]. Maize silk contains various constituents essential for human diet such as maizaniid acid, fixed oils, resin, sugar, mucilage, salt, and fibers [27].

So, this experiment was conducted with an objective to study the effect of sodium alginate at different concentrations on growth parameters, soil physical and chemical properties and some elements content during vegetative stages of *Corchorus olitorius*, *Vicia faba* and *Zea mays* plants.

2. Materials and methods

Plants used and growth conditions

Pure stains of *Zea mays* L. (maize), *Vicia faba* (bean) and *Corchorus Olitorius* (mallow) were obtained from the Research Centre of Agriculture, Agriculture Ministry, Egypt. The utilized chemicals in this investigation were of analytical grade.

Time course of experiment:

A homogenously-sized lot of *Zea mays* L. (maize), *Vicia faba* L. (bean) and *Corchorus olitorius* (mallow) seeds were selected and sterilized by soaking in 0.01% HgCl₂ solution for 3 minutes. After the seeds had been washed by distilled water, the seeds of each plant were divided into 4 equal groups each contains 100 seeds and were soaking for four hours as follow:

Group (1): Socked in water to serve as a control.

Group (2): Soaked in 25 ppm sodium alginate solution.

Group (3): Soaked in 50 ppm sodium alginate solution.

Group (4): Soaked in 100 ppm sodium alginate solution.

Seeds of the four groups were cultivated at 10/10/2018 in earthen pots (20 cm in width) containing the same soil amounts (sand: clay, 1:2 v/v). Some physical and chemical analysis were carried out to the used soil sample according to [9]. After that super phosphate fertilizer (0.5 g/pot) addition to the soil before sowing, ten seeds were sown in each pot and irrigated as usual practice by addition of the same amounts of water to each pot when required. Plants were exposed to normal day night conditions. After 10 days from sowing thinning takes places where 5 uniformed seedlings only left to grow in each pot.

Three successive sampling, during vegetative stage takes place; after 10, 20 and 40 days from sowing, representing the first, the second and the third (I, II and III) stages respectively.

The collected samples of the three stages were used for assessment of growth parameters (shoot length, root length, plant fresh & dry weights and plant water percentage)/ plant. some elements (N, P, K and Na). in addition soil physical and chemical properties were also determined but only for maize plant (after 20 days from sowing of treated and untreated plant); as this plant have the best response for alginate treatments.

Data were analyzed by least significant difference (LSD) test at probability of 0.05 to identify significant effect of a treatment. ANOVA analysis was done with the IBM SPSS-20 statics software [51]

1- Determination of growth parameters:

The collected samples of the three stages were used for assessment of growth parameters (shoot length, plant fresh & dry weights, root length and water percentage).

2-Soil sampling and analysis:

Collection of samples of soil from different pots (5-10 cm depth) representing the treated and untreated *Zea mays* after 20 days from

planting. Bring these samples to the laboratory in closed plastic bags after collection. Samples were brought to the laboratory, air dried, thoroughly mixed, passed through a 2 mm sieve to remove debris and gravel and then packed in plastic bags to be ready for physical and chemical analyses.

Physical characteristics: The heavy textured samples of soil were determined by using Bouyoucous hydrometer method as described by [44]. While the coarse textured soil samples determined as recommended by [39].

Meanwhile the available water (AW) determined according to [26] by the following formula

$$AW = FCV - PWP$$

Chemical characteristics:

The chemical variables estimated in the present study included, calcium carbonate, organic carbon, pH, electrical conductivity, chlorides, sulphates, carbonates, bicarbonates, total nitrogen, total dissolved phosphorous and extractable cations (K^+ , Ca^{++} and Mg^{++}). Calcium carbonate and organic matter were estimated by using air dry soil samples, while other variables were determined using soil water extract (1:5).

Soluble cations and anions

in the soil paste extract were determined according to methods described by [22].

Calcium and magnesium were determined by titrating with versenate (EDTA) using murexide as an indicator for calcium, eriochrome black T as an indicator for Ca^{++} and Mg^{++} . **Potassium** was determined by flame photometer. **Carbonate and bicarbonate** were determined by titration with HCl using phenolphthalein as an indicator for the former and methyl orange as an indicator for the latter. **Chloride** was determined using Mohr's method. **Sulphate** was calculated by subtracting the total soluble anions from the total soluble cations.

Soil pH was determined in the saturated soil paste using a Gillenkamp pH meter (Model pH Tester 2TM) and total soluble salts were determined by measuring the **electrical conductivity (EC)** both according to [49].

Organic matter content was determined using Walkely's rapid titration method [21].

Calcium carbonate was determined using Collin's calcimeter as described by [43].

3-Estimation of elements

Estimation of total-N:

Determination of total nitrogen by the conventional semi-micropropagation of Kjeldahl method of [48] and described by [14].

Estimation of phosphorus:

This method depends on the formation of a blue complex between phosphate and molybdic acid in presence of a reducing agent. The following method of [29] as described by [19] and adopted by [14], for compounds extraction of the total phosphorus were essentially similar to those described by [8].

Estimation of K and Na:

Wet ashing method, plant materials were dried in an oven at 80° C till constant weight. The dried matter digested according to the method of [10]. Digested the samples on electric heater until dense white fumes appeared and finally the solution after filtration became clean and was about 5 ml. The solution was stored for potassium (K) determination by the technique of the flame emission as adopted by [47], while to determine leaf content of Na^+ was determined using a flame photometer type M7D, according to [3].

3. Results and Discussion

Changes in growth parameters:

A- *Corchorus olitorius* plant:

At first vegetative stage of *Corchorus olitorius* plant growth, the obtained results in table (1) showed that, the treatment with 25 and 50 ppm sodium alginate increased the tabulated growth parameters either significantly (shoot & root lengths) or non-significantly (fresh & dry weights and water percentage/ plant). The treatment with 100 ppm sodium alginate increased shoot length significantly and fresh weight, root length & water percentage non significantly and there was nearly no change in dry weight as compared to control values.

In the second stage, a non-significant decrease in water content with the three used treatments, also, in the fresh weight affected by 100 ppm SA, the trend of changes in the other determined parameters was more or less similar to that at the first vegetative stages of

Corchorus olitorius plant, comparing to the untreated values.

At the third vegetative stages, as compared to control values, except water percentage which recorded a non-significant decrease, while, the data recorded a significant increase in the shoot length. Also a non-significant increase in the other parameters (fresh & dry weights and root length), as compared to control.

B- *Vicia faba* plant:

The obtained results in table (2) showed that, at first vegetative stage of *Vicia faba* plant growth, the treatment with 25, 50 and 100 ppm sodium alginate increased the tabulated growth parameters either significantly (shoot length) or non-significantly (fresh & dry weights, root length and percent of water content of seedling) as compared to control values.

It was recorded at second vegetative stage of faba bean, that 25 ppm sodium alginate caused a significant increase in shoot length and non-significant increase in the other determined parameters. Treatment with 50 ppm and 100 ppm sodium alginate caused a non-significant increase in dry weight and root length and significant increase in shoot length, fresh weight and water percentage as compared to control values.

In relation to control values, at the third vegetative stage the effect of 25, 50 and 100 ppm sodium alginate on *Vicia faba* plant showed a significant increase in shoot and root lengths, while a non-significant increase shown in plant fresh and dry weight. On the other hand, water percentage showed non-significant decrease as compared to control values.

C- *Zea mays* plant:

The obtained results in table (3) showed that, at first vegetative stage of *Zea mays* plant, the treatment with 25 ppm sodium alginate increased the tabulated growth parameters either significantly (fresh weight and water percentage) or non-significantly (shoot length, dry weight and root length). In corresponding to 50 ppm sodium alginate treatment, significant increases in shoot length, fresh weight, root length and water percentage of the plant was obtained. On the other hand, dry

weight increased non-significantly. The treatment with 100 ppm sodium alginate increased fresh & dry weights and root length non significantly but shoot length & water percentage increased significantly, as compared to control values.

At second vegetative stage of *Zea mays*, it was recorded that 25, 50 and 100 ppm sodium alginate caused non significant increase in fresh & dry weights and water percentage and significant increase in shoot length and root length was obtained as compared to control values.

At the third vegetative stage, 25 ppm sodium alginate caused a significant increase in root length and non-significant increase in fresh & dry weight and shoot length but decreased water percentage non significantly. The treatment with 50 ppm sodium alginate caused significant increase in shoot length, fresh weight, root length & water content and non-significant increase in dry weight. The application of 100 ppm sodium alginate showed significant increases in shoot & root lengths while, a non-significant increases in fresh & dry weights and a non significant decrease in water percentage as compared to control values. In this connection, many studies indicated that marine algae polysaccharides and derived oligosaccharides can stimulate growth in different plants [13, 56, 57, 15]

In a study, to evaluate the effect of the concentration of NaAlg submicroparticles (0–100 µg/mL) on germination, shoot & root lengths, the number of nodes & leaves, fresh & dry weight and NPK and Na contents [31]. The results had showed that NaAlg submicroparticles can exert biological activity at higher concentrations (100 µg/mL). They concluded that special properties of submicroparticles can improve the absorption and utilization in plant.

Of interest to observed that 50 ppm SA caused the best influence in most cases, in this concern, the concentration is important for its resulting effect. Algae polysaccharides can promote the elongation of roots of rice and carrot at the concentration of 500µg/mL [54]. Other study showed that at a concentration of 2×10^4 µg/mL alginate can stimulate the germination of seed and plant length of

sunflower [4]. Moreover, a previous study also showed that at a concentration of 2000 µg/mL alginate-derived oligosaccharides can promote the growth of cucumber and tomato plants [32,30]. In addition, at concentrations of 750 µg/mL and 1500 µg/mL alginate-derived oligosaccharides stimulate germination *via* the promotion of the activity and acceleration of

amylase of the metabolic activities of the maize seed [18]. It was previously reported that the high concentrations of oligosaccharides can induce cell apoptosis in plants [53, 56]. They speculated that the high concentrations of CaAlg submicroparticles may have a similar effect on plants

Table (1): Effect of different sodium alginate concentrations on growth parameters of *Corchorus olitorius* plant during three vegetative stages.

| Stages | Treatments | Parameters/ plant | | | | |
|-----------|------------|-------------------|----------------------|---------------------|-----------------|---------------------|
| | | Shoot Length(cm) | Plantfresh weight(g) | plant dry weight(g) | Root length(cm) | Water percentage(%) |
| Stage I | Control | 7.04 | 16.72 | 1.98 | 12.78 | 88.15 |
| | 25 ppm | 13.68* | 18.32 | 1.99 | 14.42 | 89.95 |
| | 50 ppm | 16.28* | 20.30 | 2.39 | 14.84 | 88.22 |
| | 100 ppm | 13.96* | 19.97 | 2.30 | 14.06 | 88.48 |
| Stage II | Control | 14.84 | 13.17 | 1.75 | 13.24 | 86.71 |
| | 25 ppm | 25.7* | 15.94 | 1.78 | 15.3 | 88.83 |
| | 50 ppm | 26.24* | 21.03* | 2.25 | 15.9 | 89.30* |
| | 100 ppm | 21.98* | 20.47* | 1.92 | 14.54 | 90.62* |
| Stage III | Control | 27.42 | 31.47 | 3.38 | 14.66 | 89.25 |
| | 25 ppm | 32.02* | 32.72 | 3.59 | 17.87* | 89.02 |
| | 50 ppm | 36.92* | 32.73 | 5.05 | 20.66* | 84.54 |
| | 100 ppm | 32.54* | 32.33 | 3.78 | 18.18* | 88.30 |

(*)=significant increase or decrease at 0.05 LSD

Table (2): Effect of different sodium alginate concentrations on growth parameters of *Vicia faba* plant during three vegetative stages. (*)=significant increase or decrease at 0.05

| Stages | Treatments | Parameters/ plant | | | | |
|-----------|------------|-------------------|-----------------------|---------------------|-----------------|---------------------|
| | | ShootLength (cm) | plant fresh weight(g) | plant dry weight(g) | Root length(cm) | Water percentage(%) |
| Stage I | Control | 5.12 | 5.49 | 0.96 | 12.28 | 82.51 |
| | 25 ppm | 6.86 | 6.47* | 0.96 | 13.5 | 85.61* |
| | 50 ppm | 8.66* | 6.59* | 0.97 | 16.08* | 85.28* |
| | 100 ppm | 7.78* | 6.36 | 0.96 | 13.44 | 84.90* |
| Stage II | Control | 7.3 | 9.71 | 1.30 | 14.78 | 86.61 |
| | 25 ppm | 9.14* | 13.86 | 1.58 | 21.78* | 88.60 |
| | 50 ppm | 10.58* | 13.97 | 1.59 | 24.46* | 88.61 |
| | 100 ppm | 9.88* | 13.91 | 1.37 | 21.90* | 90.15 |
| Stage III | Control | 15.9 | 38.25 | 4.91 | 18.22 | 87.16 |
| | 25 ppm | 21.7 | 45.34 | 8.15 | 24.02* | 82.02 |
| | 50 ppm | 29.20* | 88.67* | 9.51 | 35.56* | 89.27* |
| | 100 ppm | 21.94* | 51.96 | 7.56 | 27.24* | 85.45 |

Table (3): Effect of different sodium alginate concentrations on growth parameters of *Zea mays* plant during three vegetative stages.

| Stages | Treatments | Parameters/ plant | | | | |
|-----------|------------|-------------------|-----------------------|---------------------|------------------|----------------------|
| | | Shoot Length(cm) | plant fresh weight(g) | plant dry weight(g) | Root length (cm) | Water percentage (%) |
| Stage I | Control | 5.12 | 5.49 | 0.96 | 12.28 | 82.51 |
| | 25 ppm | 6.86 | 6.47* | 0.96 | 13.5 | 85.61* |
| | 50 ppm | 8.66* | 6.59* | 0.97 | 16.08* | 85.28* |
| | 100 ppm | 7.78* | 6.36 | 0.96 | 13.44 | 84.90* |
| Stage II | Control | 7.3 | 9.71 | 1.30 | 14.78 | 86.61 |
| | 25 ppm | 9.14* | 13.86 | 1.58 | 21.78* | 88.60 |
| | 50 ppm | 10.58* | 13.97 | 1.59 | 24.46* | 88.61 |
| | 100 ppm | 9.88* | 13.91 | 1.37 | 21.90* | 90.15 |
| Stage III | Control | 15.9 | 38.25 | 4.91 | 18.22 | 87.16 |
| | 25 ppm | 21.7 | 45.34 | 8.15 | 24.02* | 82.02 |
| | 50 ppm | 29.20* | 88.67* | 9.51 | 35.56* | 89.27* |
| | 100 ppm | 21.94* | 51.96 | 7.56 | 27.24* | 85.45 |

Table (4) : Effect of different concentrations of sodium alginate on the nitrogen , phosphorus, potassium and sodium percentages (%)of *Corchorus olitorius*, *Vicia faba* and *Zea mays* through the second vegetative stage (after 20 days from sowing)

| Treatment | | Parameters | | | |
|----------------------------|---------|------------|-------|-------|--------|
| | | N% | P% | K% | Na% |
| <i>Corchorus olitorius</i> | Control | 25.2 | 1.89 | 1.14 | 0.092 |
| | 25 ppm | 30.5* | 2.68* | 2.60* | 0.095* |
| | 50 ppm | 35.7* | 2.29* | 3.10* | 0.100* |
| | 100 ppm | 18.3* | 1.37* | 3.00* | 0.120* |
| <i>Vicia faba</i> | Control | 26.1 | 1.95 | 0.70 | 0.149 |
| | 25 ppm | 33.9* | 2.54* | 0.71 | 0.152* |
| | 50 ppm | 47.0* | 3.41* | 0.73* | 0.160* |
| | 100 ppm | 46.1* | 3.46* | 0.71 | 0.181* |
| <i>Zea mays</i> | Control | 62.6 | 2.7 | 0.25 | 0.101 |
| | 25 ppm | 42.6* | 3.19* | 1.02* | 0.195* |
| | 50 ppm | 41.2* | 3.21* | 1.29* | 0.152* |
| | 100 ppm | 40.9* | 3.07* | 1.16* | 0.166* |

(*) = significant increase or decrease at 0.05 LSD

Changes in some elements content:

A - *Corchorus olitorius* plant:

The data in table (4) revealed that, treatment with 25 and 50 ppm sodium alginate significant increase nitrogen, phosphorus, potassium and sodium content. A marked significant increment in K and Na content was detected in the *Corchorus olitorius* plant, at the second vegetative stage by application of 100 ppm sodium alginate, meanwhile N and P showed a significant decrease as compared to control values.

B- *Vicia faba* plant:

According to the data in table (4), the treatment with 25, 50 and 100 ppm sodium alginate increased nitrogen, phosphorus and sodium content significantly, meanwhile K showed a non-significant increase at 25 and 100 ppm but a significant increase at 50 ppm, as compared to control values.

C- *Zea mays* plant:

Perusal of the data in table (4) revealed that, except for treatment with 25, 50 and 100 ppm sodium alginate which decreased nitrogen content significantly, a marked significant increment in P, K and Na content was

detected in the *Zea mays* plant, as compared to the corresponding values of the untreated plants.

In this regard, absorption and utilization of mineral nutrients facilitated by irradiated SA [23,46]. Similar stimulation in leaf N, P and K contents due application of irradiated SA to various other crops has been reported by [23, 25]. Application of different concentrations of irradiated SA on the *Papaver somniferum L.* leaves and reported that at 120 ppm concentration of irradiated SA improved leaf nitrogen content to the maximum [2]. Compared to control, the Na concentration in roots of plants exposed to CeO₂ NPs alone decreased by 10.5%. In the CeO₂ NP

alginate treatments, Na in roots reduced up to 16%. This could be associated with the increase in K absorption. As a C₄ plant, corn needs Na for phosphoenolpyruvate regeneration, the carboxylation substrate [52, 58]. [35] study the influence of irradiated sodium alginate (ISA) on *foeniculum vulgare* and found that leaf N, P &K contents increased significantly as a result of application of ISA.

Polysaccharide submicroparticles have been applied in the agricultural field, such as the incorporation of (NPK) fertilizer into the submicroparticles to save fertilizer consumption. [11]. A research data [38] reported that at higher concentrations of

alginate-derived oligosaccharides exerts its biological functions Furthermore

,submicroparticles have a nanometer size effect, which can stimulate its bioactivities

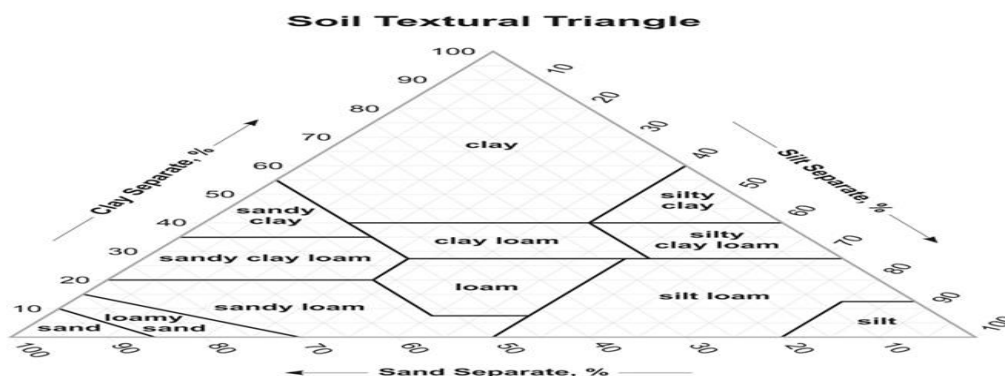
.Changes in soil physical and chemical properties:

As mentioned before, the soil properties were carried out only in the soil where the treated and untreated *Zea mays* plant sowing at stage II.

Table 5 indicated that, the clay, silt and porosity percent increased by treatment with 25,50 and 100 ppm sodium alginate except for porosity% which decreased by treatment with 25 ppm sodium alginate when compared with control. Meanwhile the sand percent decreased as compared to control values. In soils planted with that treated by 50 and 100 ppm sodium alginate the field capacity percentage (F.C.P) and wilting point (W.P) percents increased whereas no change detected at 25 ppm sodium alginate as compared to control values. The soil texture not changed by treatment with 25 ppm sodium alginate but changed from sandy loam (in the untreated plants) into loamy soil at 50 ppm sodium alginate and to sandy clay loam by application of 100 ppm sodium alginate; according to [44] as illustrated in table 5 & fig(1).

Table (5) The physical properties for the planting soils of *Zea mays* plant after 20 days from sowing.

| Parameters Treatment | Sand% | Clay% | Silt% | Prosity% | F.C.P% | W.P% | Soil texture |
|----------------------|-------|-------|-------|----------|--------|-------|-----------------|
| Control | 62.16 | 16.05 | 19.44 | 17.91 | 18.00 | 36.00 | Sandy loam |
| 25 ppm | 57.99 | 16.17 | 23.73 | 17.38 | 18.00 | 36.00 | Sandy loam |
| 50 ppm | 48.83 | 18.35 | 36.00 | 23.00 | 28.00 | 56.00 | Loamy |
| 100 ppm | 52.90 | 20.18 | 22.80 | 19.60 | 27.00 | 54.00 | Sandy clay loam |



F.C.P: field capacity percentage W.P: wilting point

Fig. (1): Soil texture pyramid [44]

Chemical properties, in table 6 detected increases in calcium carbonate & organic matter and potassium cations in response to treatment with 25, 50 and 100 ppm sodium alginate, while there was a decrease in nitrogen and phosphorus contents, as compared to control values.

According to table (6), as compared to control values, application of 25, 50 and 100 ppm sodium alginate caused increases in EC, (HCO_3^- , SO_4^{--}) anions ,(K^+ , Na^+ , Ca^{++}) cations and T.S.S ,whereas Cl^- anion increased only at 50 and 100 ppm sodium alginate but decreased at 25 ppm sodium alginate. Meanwhile there was a decrease in pH and Mg^{++} cations by application of 25, 50 and 100 ppm sodium alginate. In this

connection, it was recorded that sodium alginate in smallest amounts brings about relatively the greatest increments in the contents of water of the soil at which the aeration factor (A.F). Also in the alginate treated soils the water-holding power and pH increased. As the beginning pH (7.3) of the medium and the final pH. (7.7) at the termination of the experiments, were unaltered by the presence of the sodium alginate. Hence it may be concluded that the changes of hydrogen-ion concentration in the soil brought about by sodium alginate addition cannot account for, and may not even be contributing to, the increased A.F. [45]. They suggested that the results make it evident that the change in A.F. value with increase in water content is a function of both water-holding power and crumb stability.

Moreover, the experimental results showed that as sodium alginate in the soil increased, the liquid limit and plastic limit of the soil also increased. The above results indicated that the soil properties in both the physical structure and chemical composition can be changed significantly by using sodium alginate. Large amount of $-\text{COO}-$ groups included in Sodium alginate. Sodium alginate when dissolved in water giving rise to adhesion and it becomes polyanionic. Under mild conditions, sodium alginate can combine quickly with water to form a hydrogel [42, 59].

Table (6): The chemical properties for the planting soils of *Zea mays* after 20 days from sowing.

| Treatments | CaCO ₃ % | Organic matter % | N ppm% | P ppm % | K ppm% |
|------------|------------------------|---------------------|-----------|------------|-----------|
| Control | 0.85 | 0.61 | 661.50 | 15.10 | 20.67 |
| 25 ppm | 0.95 | 1.14 | 581.00 | 14.30 | 20.72 |
| 50 ppm | 1.40 | 2.01 | 487.00 | 12.83 | 21.02 |
| 100 ppm | 1.60 | 2.52 | 395.50 | 8.92 | 22.16 |

Table (7): The chemical properties for the planting soils of *Zea mays* plant after 20 days from sowing.

| Treatments | CaC % | Organic matter % | N ppm% | P ppm % | K pp% |
|------------|----------|---------------------|-----------|------------|----------|
| Control | 0.85 | 0.61 | 661.50 | 15.10 | 20.67 |
| 25 ppm | 0.95 | 1.14 | 581.00 | 14.30 | 20.72 |
| 50 ppm | 1.40 | 2.01 | 487.00 | 12.83 | 21.02 |
| 100 ppm | 1.60 | 2.52 | 395.50 | 8.92 | 22.16 |

Conclusion

In conclusion, the result of this study showed that the application of SA especially 50 ppm had a promotive effect on plant growth by enhancing growth parameters, elements absorption and changing soil physical and chemical properties.

4. References

1. Aftab, T.; Khan, M.M.A.; Idrees; M.; Naeem, M.; Moinuddin.; Hashmi N. and Varshney L. (2011): Enhancing the growth, photosynthetic capacity and artemisinin content in *Artemisia annua L.* by irradiated sodium alginate. *Radiat. Phys. Chem.* (2011), **80**: 833-836.
2. Aktar, F.; Khan, M.A. and Mustafa, A.I. (2017): Effect of Gamma Irradiated Sodium Alginate on Selective Crops as Plant Growth Promoter and Bug control action. *Int J Nano Med & Eng.* **2:10**,163-172
3. Allen, L.M. and Richard, F.S. (1986): Determination of Ca, Mg, Na, Cd, Cu, Fe, K, Li and Zn in acid mine and reference water samples by inductively coupled plasma atomic fluorescence spectrometry, *Analyst*,**111**, 645-649
4. Anis, M.; Zaki, M.J.; Dawar, S. (2012): Development of a Na-alginate-based bioformulation and its use in the management of charcoal rot of sunflower

- (*Helianthus annuus L.*). *Pak. J. Bot.*, **44**, 1167–1170.
5. Anthony, J.; Gabapathy, A.; Coothan, K. V.; Streenivasan, P. P.; Arjuna, R. and Palaninathan, V. (2007): Beneficial effects of sulphated polysaccharides from *Saragassum wightii* against mitochondrial alterations induced by cyclosporine A in rat kidney *Molecular Nutrition and Food Research*, **51**, 1413-1422.
 6. Attere, F. (1999): Note introductive de l'atelier sur les légumes feuilles. In: Chwey, J.A., Eyzaguire, P. (Eds.), *Biodiversity of Traditional Leafy Vegetables in Africa*. International plant Genetic Resources Institute (IPGRI) via delle sette chiese 14200145, Rome, Italie 150p.
 7. Bailey, J.M. (2003): *Aliments du Pacifique: les feuilles vertes que nous mangeons*. Version française du manuel de la CPS n° 31, 2000. Service de publication du Secrétariat général de la communauté du pacifique (CPS), Graphoprint, Nouméa 97p.
 8. Barker, J. and Mapson, L.W. (1964): Studies on the respiratory and carbohydrate metabolism of *plant tissues*. *J. of Exp. Bot.*, **15**: 272- 283.
 9. Black, C.A. (1965): *Methods of soil analysis. Part I. Physical and mineralogical properties. Inducing statistics of measurement and sampling*. American Soc. Agronomy, Inc. Madison, Wisconsin.
 10. Chapman, H.D. and Pratt, P.F. (1978): *Method of Analysis for Soil, Plant and Water*, California University, Division Agrc. Sci., Priced Publication, pp 50,169.
 11. Corradini, E.; de Moura, M.R.; Mattoso, L.H.C. (2010): A preliminary study of the incorporation of NPK fertilizer into chitosan nanoparticles. *Express. Polym. Lett.*, **4**, 509–515.
 12. CRA. (2006): *Corn oil* (5th ed.). Washington, DC: Corn Refiners Association.
 13. González, A.; Castro, J.; Vera, J.; Moenne, A. (2012): Seaweed oligosaccharides stimulate plant growth by enhancing carbon and nitrogen assimilation, basal metabolism, and *cell division*. *J. Plant Growth Regul.*, **32**, 443–448.
 14. Haroun, S.A. (1985): Studies on adaptation of plants to water stress. Ph. D. Thesis, Bot. Department, Fac. Sci. Mans. Univ. Mansoura. Egypt.
 15. He, X.Y., Kong, Q.Q. and Xi, X.M. (2008): Security testing of 20% kefucuo seed coating agent on Seeds. *Inner Mongolia Agricul. Sci. Technol.*, **5**: 60-61.
 16. Hegazy, E.A., Abdel-Rehim, H.A., Diao, D.A., & El-Barbary, A. (2009): Report: Controlling of degradation effects in radiation processing of polymers. In: *Controlling of Degradation Effects in Radiation Processing of Polymers*. International Atomic Energy Agency. Vienna, Austria. 2009, pp. 64-84.
 17. Hien, N.Q.; Nagasawa, N.; Tham, L.X.; Yoshii, F.; Dang, H.V., Mitomo, H.; Makuuchi, K. and Kume, T. (2000): Growth promotion of plants with depolymerised alginates by irradiation. *Radiat. Phys. Chem.*, **59**: 97-101
 18. Hu, X.; Jiang, X.; Hwang, H.; Liu, S. and Guan, H. (2004): Promotive effects of alginate-derived oligosaccharide on maize seed *germination*. *J. App. Phycol.* **2004**, **16**: 73–76.
 19. Humphries, E.C. (1956): Mineral components and ash analysis. In *modern method of plant analysis*. Vol. **1**, (edited by K. Paech and M.V. Tracey). 468–502. Berlin: Springer Verlag.
 20. Idrees, M.; Naeem, M.; Alam, M.; Aftab, T.; Hashmi, N.; Khan, M.M.A.; Moinuddin and Varshney, L.(2011): Utilizing the gamma irradiated sodium alginate as a plant growth promoter for enhancing the growth, physiological activities and alkaloids production in *Catharanthus roseus L.* *Agric. Sci. China*, **10**, 1213-1221.
 21. Jackson, M.L. (1962): *Soil chemical analysis*. Constable Co. Ltd. London, p. 296.
 22. Jackson, M.L. (1973): *Soil chemical analysis*. Prentice-Hall Inc., Englewood Cliffs, New Jersey, USA, 225.
 23. Jamsheer, M.K. (2010): Response of beet root (*Beta vulgaris L.*) to the application of phosphorus and gamma-irradiated

- sodium alginate. M. Sc. Thesis. Aligarh Muslim University. India.
24. Kar, C.S., Kundu, A., Sarkar, D., Sinha, M.K., Mahapatra, B.S., (2009): Genetic diversity in jute (*Corchorus* spp) and its utilization: a review. *Indian J. Agric. Sci.* **79**, 575–585.
 25. Khan Z.H.; Khan MMA, Aftab T.; Idrees, M. and Naeem M (2010): Influence of alginate oligosaccharides on the growth, yield and alkaloids production of opium poppy (*Papaver somniferum*L.). *Front Agric China* (DOI: 10. 1007/s11703-010-1056-0).
 26. Kirkham, M. B. (2005): Principles of soil and plant water relations. Elsevier Academic Press.
 27. Kumar, D., & Jhariya, N. A. (2013): Nutritional, medicinal and economical importance of corn: A mini review. *Research Journal of Pharmaceutical Sciences*, **2**, 7–8.
 28. Kume, T.; Nagasawa, N. and Yoshii, F. (2002): Utilization of carbohydrates by radiation processing. *Radiat. Phys. Chem.*, **63**: 625627
 29. Kuttner, T. and Lichtenstein, L. (1932): Micro-colorimetric studies: Estimation of organically-bound phosphorus. A system of analysis of phosphorus compounds in blood. *J. of biological chem.*, **95**: 661-670.
 30. Li, J.; Wang, X.; Lin, X.; Yan, G.; Liu, L.; Zheng, H.; Zhao, B.; Tang, J. and Guo, Y.D. (2018): Alginate-derived oligosaccharides promote water stress tolerance in cucumber (*Cucumis sativus* L.). *Plant Physiol. Biochem.*, **130**, 80–88.
 31. Liang, J.; Yan, H.; Puligundla, P.; Gao, X.; Zhou, Y.; Wan, X. (2017): Applications of chitosan nanoparticles to enhance absorption and bioavailability of tea polyphenols: A review. *Food Hydrocoll*, **69**, 286–292
 32. Liu, R.; Jiang, X.; Guan, H.; Li, X.; Du, Y.; Wang, P.; Mou, H. (2009): Promotive effects of alginate-derived oligosaccharides on the inducing drought resistance of tomato. *J. Ocean Univ. China*, **8**, 303–311.
 33. Macarulla, M. T.; Medina, C.; Diego, M.; Chavarri, M.; Zulet, M.; Mart´inez J. A.; N´oel-Suberville, C.; Higuere P. and Portillo, M. P. (2001): Effects of the whole seed and a protein isolate of faba bean (*Vicia faba*) on the cholesterol metabolism of hyper cholesterolaemic rats. *Br J Nutr* **85(05)**:607–614.
 34. Mateos-Aparicio, I.; Redondo-Cuenca, A.; Villanueva-Su´arez, M.; Zapata-Revilla, M. and Tenorio-Sanz, M.(2010): Pea pod, broad bean pod and okara, potential sources of functional compounds. *LWT – Food Sci Technol* **43(9)**:1467–1470.
 35. Nadeem, H. (2013): influence of gamma irradiated sodium alginate and some macronutrients on the essential oil production in *Foeniculum Vulgar* mill, Department of Botany, Aligarh Muslim University, Aligarh (u.p) India
 36. Nafziger, E.D. (1996): Effect of Missing and Two-Plant Hills on Corn Grain Yield. *Journal of Production of Agriculture*, **9**: 238-240 [Research report detailing different effects of missing or doubled maize plants on yields].
 37. Ngomuo, M.S., Stoilova, T., Feyissa, T., Kassim, N., Ndakidemi, P.A., (2017): Leaf and seed yield of jute mallow (*Corchorus olitorius* L.) accessions under field conditions for two consecutive growing seasons. *J. Hortic. Sci. Biotechnol.* **92**, 614–620
 38. Nguyen Van, S.; Dinh Minh, H. and Nguyen Anh, D. (2013): Study on chitosan nanoparticles on biophysical characteristics and growth of Robusta coffee in green house. *Biocatal. Agric. Biotechnol.*, **2**, 289–294.
 39. Obi, M.A. (1974): The wilting point and available moisture of tropical forest soils in Nigeria. *Exp.Agric.*10:305-312.
 40. Ofuya, Z. M. and Akhidue, V. (2006): The role of pulses in human nutrition: a review. *J Appl Sci Environ Manag* **9(3)**:99–104.
 41. Orthoefer, F., Eastman, J., & List, G. (2003): Corn oil: composition, processing and utilization. In P. J. White, L. A. Johnson (Eds.), *Corn: Chemistry and technology* (2nd ed., pp. 671–693). St. Paul, MN: American Association of Cereal Chemists

42. Ouwerx, C.; Velings, N. and Mestdagh M. (1998): Physico-chemical properties and rheology of alginate gel beads formed with various divalent cations. *Polym Gels Netw* 6:393–408.
43. Page, A.L.; Miller, R.H. and Keeney, D.R. (1982): *Methods of soil analysis, part2. Chemical and microbiological properties*, American Society of Agronomy, Inc., Publisher, Madison, Wisconsin USA, 2-13.
44. Piper, C.S. (1947): *Soil and plant analysis*. Interscience Publishers, Inc. New York.
45. Quastel, J.H.; Vebley, D.M. (1947): the effects of the addition to soil of alginic acid and of other forms of organic matter on soil aeration, *J. Agric. Sci.* **37**, 249.
46. Qureshi, A.H. (2010): Effect of nitrogen and gamma-irradiated sodium alginate on the efficiency of beetroot (*Beta vulgaris L.*). M.Sc. Thesis. Aligarh Muslim University. India.
47. Ranganna, S. (1977): *Manual Analysis of fruit and vegetable products*. Tata MC craw-Hill publishing company limited New York.
48. Rees, M.W. and Williams, E.F. (1943): The total nitrogen content of egg albumin and other proteins. *Journal of Biochemistry*, **37**: 354-359.
49. Richards, L.A. (1954): *Diagnosis and improvement of saline and alkali soil*, United states department of Agriculture, hand book60-160.
50. Singh, A. K.; Bharati, R. C.; Manibhushan, N. C. and Pedpati, A. (2013): An assessment of faba bean (*Vicia faba L.*) current status and future prospect. *Afr J Agric Res* **8(50)**:6634–41
51. Snedecor, G.W. and Cochran, W.G. (1982): *Statistical Methods*. The owa State University Press 7th Edit, 2nd Printing. 507 pp. *Soilm Sci.*, **81**: 173–192.
52. Taiz, L., Zeiger, E., (1998): *Plant Physiology*, 2nd ed. Sinauer, Sunderland, Massachusetts
53. Wang, W.X.; Li, S.G.; Zhao, X.M.; Du, Y.G.; Lin, B.C. (2008): Oligochitosan induces cell death and hydrogen peroxide accumulation in tobacco suspension cells. *Pestic. Biochem. Physiol.*, **90**, 106–113.
54. Xu, X.; Iwamoto, Y.; Kitamura, Y.; Oda, T.; Muramatsu, T. (2003): Root growth-promoting activity of unsaturated oligomeric uronates from alginate on carrot and rice plants. *Biosci. Biotechnol. Biochem.*, **67**, 2022–2025.
55. Zhang, H.Y.; Wang, W.X.; Yin, H.; Zhao, X.M.; Du, Y.G. (2012): Oligochitosan induces programmed cell death in tobacco suspension cells
56. . Zhang, Y.; Liu, H.; Yin, H.; Wang, W.; Zhao, X.; Du, Y. (2013): Nitric oxide mediates alginate oligo saccharides-induced root development in wheat (*Triticum aestivum L.*). *Plant Physiol. Biochem.*, **71**, 49–56.
57. Zhang, Y.; Yin, H.; Zhao, X.; Wang, W.; Du, Y.; He, A.; Sun, K. (2014): The promoting effects of alginate oligosaccharides on root development in *Oryza sativa L.* mediated by auxin signaling. *Carbohydr. Polym.*, **113**, 446–454.
58. Zhao, L.; Videa, J.R.P.; Peng, B.; Bandyopadhyay, S.; Diaz, B.C.; Avila, P.O.; Montes, M.O; Keller, A.A.; Torresdey, J.L.G. (2014): Alginate modifies the physiological impact of CeO₂ nano particles in corn seedlings cultivated in soil. *Journal of Environmental Sciences* **26** 382–389
59. Zhao, Y.; Zhuang, J.; Wang, Y.; Jia, Y.; Niu, P.; Jia, K. (2019): Improvement of loess characteristics using sodium alginate, Springer-Verlag GmbH Germany, part of Springer Nature, *Bulletin of engineering Geology and the environment*